

PRODUCT INFORMATION SHEET
Monoclonal antibodies detecting human antigens

CD21

| | | | | | |
|------|---|---|----------|---|-----------|
| PURE |  |  | IQP-109P |  | 100 tests |
| FITC |  |  | IQP-109F |  | 100 tests |
| R-PE |  |  | IQP-109R |  | 100 tests |

 **For Research Use Only**



Description

Clone B-ly4

Isotype Murine IgG1

Specificity Clone B-ly4, produces mouse IgG1 immunoglobulins directed against a 145 kD polypeptide of the human C3d receptor. Clone B-ly4 was clustered at the Leucocyte Typing Workshop IV [5].

Antigen distribution

The CD21 antigen is expressed on mature B cells, follicular dendritic cells and pharyngeal and cervical epithelial cells [1,2]. CD21, also known as the Complement receptor type 2 (CR2) or Epstein-Barr virus (EBV) receptor, is a receptor for the C3 activation fragments iC3b and C3d. CD21 antibodies have been shown to bind CD23 and IFN- α . CD21 is part of a multimeric complex on B cells which includes CD19 and CD81 [3].

Summary

The ligand for CD21 is generated during activation of complement by the attachment of C3d to potential antigen. This creates fusion proteins that may cross-link the CD19-CD21-CD81 complex to sIg on antigen-specific B cells. The resulting signal transduction lowers the threshold for cellular activation. This enables an antibody reaction to natural protein antigens containing only one or a few epitopes and therefore unable to effectively cross-link Ig molecules on specific B cells. For example, keyhole limpet hemocyanin (KLH) immune complexes can be taken up and processed by all CR2-bearing B cells, regardless of their antigen specificity. KLH immune complexes taken up this way by non-specific B cells were shown to be presented to KLH-specific, MHC class II Th cells. CD23 may also serve as a ligand for CD21 [3].

Applications

Monoclonal antibody CD21, clone B-ly4 can be applied in flow cytometry for analysis of blood and bone marrow samples or in immunohistochemistry on frozen or paraffin embedded tissue sections.

CD21 is detected by flow cytometric analysis of peripheral blood on all mature circulating (m+g+) B cells. Low density CR2 has been reported on granulocytes and monocytes, but is not detectable on normal cells by flow cytometric analysis using this monoclonal antibody. CD21 antigen has been detected very weakly on T cells. B-ly4 is applied in the immunophenotyping of leukemias and lymphomas. It is also suitable for use in immunohistochemistry for the detection of follicular dendritic cells and B cell lymphomas. In functional studies B-ly4 is suitable for studies of Epstein-Barr virus receptor and the biological function of complement components [4].

Usage

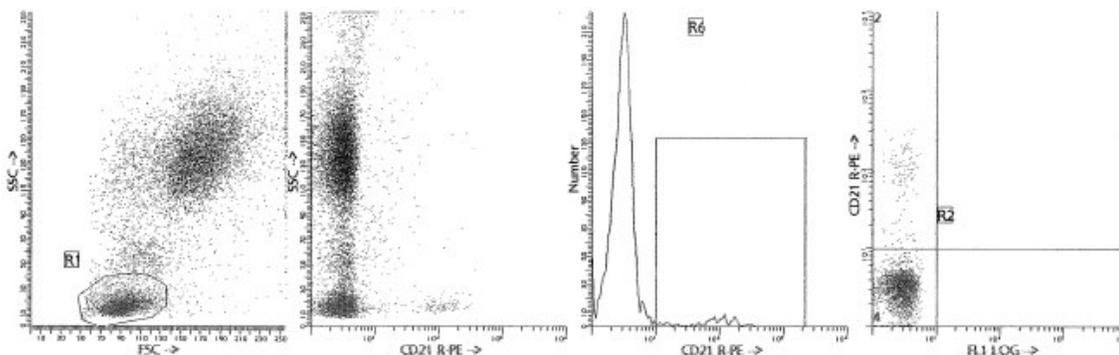
All these reagents are effectively formulated for direct immunofluorescent staining of human tissue for flow cytometric analysis using 10 μ l/10⁶ leukocytes for singles and 20 μ l/10⁶ leukocytes in case of dual and triple combinations. Since applications vary, each investigator should titrate the reagent to obtain optimal results.

HLDA Workshop

4th Leucocyte Typing Workshop - Knapp, W., et al., eds. (1990).

Representative Data

Clone B-ly4 (CD21) was analyzed by flow cytometry using a blood sample obtained from a healthy volunteer. Direct staining was performed using 10 µl of the R-PE-conjugated monoclonal antibody and 100 µl of blood sample.



Limitations

1. Conjugates with brighter fluorochromes, like PE and APC, will have a greater separation than those with dyes like FITC and CyQ. When populations overlap, the percentage of positive cells using a selected marker can be affected by the choice of fluorescent label.
2. Use of monoclonal antibodies in patient treatment can interfere with antigen target recognition by this reagent. This should be taken into account when samples are analyzed from patients treated in this fashion. IQ Products has not characterized the effect of the presence of therapeutic antibodies on the performance of this reagent.
3. Reagents can be used in different combinations, therefore laboratories need to become familiar performance characteristics of each antibody in relation with the combined markers in normal and abnormal samples.
4. Reagent data performance is based on EDTA-treated blood. Reagent performance can be affected by the use of other anticoagulants.

Reagents and materials required but not supplied

1. Flow cytometer
2. Flow cytometry disposable 12 x 75-mm capped polystyrene test tubes
3. Micropipette with disposable tips
4. Vortex mixer
5. Centrifuge
6. IQ Lyse - erythrocyte lysing solution (IQP-199)
7. IQ Starfiqs - fixation and permeabilization solution (IQP-200)
8. PBS (phosphate-buffered saline)
9. 1% paraformaldehyde solution in PBS (store at 2-8 °C in amber glass for up to 1 week)

Immunofluorescence staining and lysing protocol

- A - Flow cytometry method for use with purified monoclonal antibodies

1. Add 100 µl of EDTA-treated blood (i.e. approx. 10^6 leukocytes) to a 5 ml reagent tube. The content of one tube is sufficient to perform one test.
2. Add to each tube 10 µl of purified monoclonal antibody*. Vortex the tube to ensure thorough mixing of antibody and cells.
3. Incubate the tube for 15 minutes at room temperature in the dark.
4. Wash the labeled cells by adding 2 ml of PBS containing 0.001% (v/v) Heparin, vortexing and centrifuging (2 min 1000 x g.) and discard the supernatant.
5. Add 50 µl of 1:10 dilution of IQ Products F(ab)₂ Rabbit Anti Mouse IgG fluorescent conjugate, [FITC (IQP-190F); R-PE (IQP-190R)] in PBS containing 0.001% (v/v) Heparin to the tube. It is recommended that the tube is protected from light.
6. Mix by vortexing and incubate for 15 minutes at room temperature in the dark.
7. Add 100 µl of IQ Lyse (IQP-199 ready-to-use) and mix immediately.
8. Incubate for 10 minutes at room temperature in the dark.
9. Add 2 ml of demineralized water and incubate for 10 minutes in the dark.
10. Centrifuge the labeled cell suspension for 2 minutes at 1000 x g.
11. Remove the supernatant and resuspend the cells in 200 µl of PBS**.
12. Analyze by flow cytometry within four hours (alternatively, the cells may be fixed by 0.05% of formaline in buffered saline for analysis the next day. Some antigens are readily destroyed upon fixation and this should be taken into account when using this alternative).

- B - Flow cytometry method for use with labeled (FITC, R-PE, Cy-Q or APC) monoclonal antibodies

1. Add 100 µl of EDTA-treated blood (i.e. approx. 10^6 leukocytes) to a 5 ml reagent tube. The content of one tube is sufficient to perform one test.
2. Add to each tube 10 µl of labeled monoclonal antibody*. Vortex the tube to ensure thorough mixing of antibody and cells.
3. Incubate the tube for 15 minutes at room temperature in the dark.
4. Add 100 µl of IQ Lyse (IQP-199 ready-to-use) and mix immediately.
5. Incubate for 10 minutes at room temperature in the dark.
6. Add 2 ml of demineralized water and incubate for 10 minutes in the dark.
7. Centrifuge the labeled cell suspension for 2 minutes at 1000 x g.
8. Remove the supernatant and resuspend the cells in 200 µl of PBS**.
9. Analyze by flow cytometry within four hours (alternatively, the cells may be fixed by 0.05% of formaline in buffered saline for analysis the next day. Some antigens are readily destroyed upon fixation and this should be taken into account when using this alternative).

- C - Flow cytometry method for use with dual and triple combinations

1. Add 100 µl of EDTA-treated blood (i.e. approx. 10^6 leukocytes) to a 5 ml reagent tube. The content of one tube is sufficient to perform one test.
For combinations with anti-kappa and/or anti-lambda Ig see application note below.
2. Add to each tube 20 µl of labeled monoclonal antibody combination*.
3. Vortex the tube to ensure thorough mixing of antibody and cells.
4. Incubate the tube for 15 minutes at room temperature in the dark.
5. Add 100 µl of IQ Lyse (IQP-199 ready-to-use) and mix immediately.
6. Incubate for 10 minutes at room temperature in the dark.
7. Add 2 ml of demineralized water and incubate for 10 minutes in the dark.
8. Centrifuge the labeled cell suspension for 2 minutes at 1000 x g.
9. Remove the supernatant and resuspend the cells in 200 µl of PBS**.
10. Analyze by flow cytometry within four hours (alternatively, the cells may be fixed by 0.05% of formaline in buffered saline for analysis the next day. Some antigens are readily destroyed upon fixation and this should be taken into account when using this alternative).

* Appropriate mouse Ig isotype control samples should always be included in any labeling study

** PBS: Phosphate Buffered Saline, pH 7.2

Application note for anti-kappa and/or anti-lambda Ig combinations

Add 2 ml of PBS containing 0.001% (v/v) Heparin (**prewarmed to 37 °C**) to the cell suspension
Vortex, centrifuge (2 min at 300x g) and discard the supernatant
Repeat this step twice
Resuspend the pelleted blood cells in 100 µl PBS containing 0.001% (v/v) Heparin



Handling and Storage

Antibodies are supplied either as 100 tests per vial (1 ml) for singles or 50 tests per vial (1 ml) for dual and triple combinations. They are supplied in 0.01 M sodium phosphate, 0.15 M NaCl; pH 7.3, 0.2% BSA, 0.09% sodiumazide (NaN_3). Store the vials at 2-8 °C. Monoclonal antibodies should be protected from prolonged exposure to light. Reagents are stable for the period shown on the vial label when stored properly.

Warranty Products sold hereunder are warranted only to conform to the quantity and contents stated on the label at the time of delivery to the customer. There are no warranties, expressed or implied, which extend beyond the description on the label of the product. IQ Products is not liable for property damage, personal injury, or economic loss caused by the product.

Characterization

To ensure consistently high-quality reagents, each batch of monoclonal antibody is tested for conformance with characteristics of a standard reagent. Representative flow cytometric data is included in this data sheet.

Warning All products contain sodiumazide. This chemical is poisonous and hazardous. Handling should be done by trained staff only.

References

1. Barclay, A.N., et al. 1997. The leucocyte antigen FactsBook. Academic Press. London. 183-185
 2. Ahearn, J.M., and Fearon, D.T., 1989. Adv. Immunol. 46. 183-219
 3. Doody, G.M., et al 1996. Curr.Opin.Immunol. 8. 378-382
 4. Timens, et al. 1991. Histochemistry. 95. 605-611
 5. Knapp, W., et al., eds. 1990 Leucocyte Typing Workshop IV
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Explanation of used symbols

| | |
|---|---|
|  | Consult instructions for use |
|  | Catalogue number |
|  | Sufficient for |
|  | In Vitro Diagnostic medical device |
|  | Caution, consult accompanying document |
|  | Keep away from (sun)light |
|  | Biological risks |
|  | Temperature limitation (°C) |
|  | For Research Use Only |
|  | Batch code |
|  | Use by yyyy-mm-dd |
|  | Manufacturer |
|  | Authorized Representative in the European Community |
|  | Conformité Européenne (European Conformity) |

| | | Label - tandem | Ex -max (nm) | Em -max (nm) |
|-----|-------------|-----------------------|---------------------|---------------------|
| P | PURE | purified material | - | - |
| F | FITC | FITC | 488 | 519 |
| R | R-PE | PE | 488, 532 | 578 |
| C | CyQ | PE-Cy5.18 | 488, 532 | 667 |
| A | APC | | 595, 633, 635, 647 | 660 |
| PC | PerCP | | 488, 532 | 678 |
| PCC | PerCP-Cy5.5 | | 488, 532 | 695 |

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