

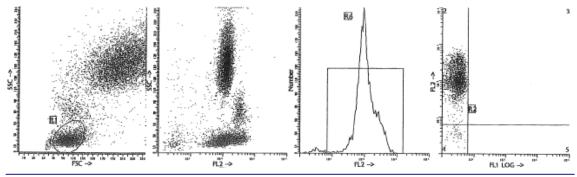
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PRODUCT INFORMATION SHEET

Monoclonal antibodies detecting human antigens

CD11a PURE R-PE	RUOREFIQP-151P▼100 testsRUOREFIQP-151R▼100 tests					
RUO For Rese	earch Use Only					
<u>íi</u>	Description					
Clone	DF1524					
Isotype	Murine IgG2b					
Specificity	Clone DF1524 produces IgG2b mouse immunoglobulins specific for CD11a. It can be used for flow cytometry and immunohistochemistry for the detection of CD11a positive leucocytes. Clone DF1524 was clustered at the Leucocyte Typing Workshop IV [5].					
Antigen distri	ribution CD11a antigens are expressed on lymphocytes, granulocytes, monocytes and macrophages, with increased levels on memory T cells [1,2]. CD11a (integrinaL subunit) combines with CD18 (integrin b2 subunit) to form the integrin LFA-1 (CD11a/CD18). The three ligands for LFA-1 are CD54 (ICAM-1), CD102 (ICAM-2) and CD50 (ICAM-3), each of which contains IgSF domains.					
Summary	CD11a/CD18 was first described as an accessory molecule in cytotoxic lymphocyte killing. LFA-1 mediates adhesion of lymphoid cells to the vascular endothelium in association with its ligands. The avidity of CD11a/CD18 to its ligands is transiently upregulated on T cells upon acitvation [3]. CD11a/CD18 has also been shown to bind bacterial lipopolysaccharides [4].					
Applications	Monoclonal antibodies CD11a, clone DF1524 can be applied in flow cytometry for analysis of blood and bone marrow samples, or in immunohistochemistry using cytospots or frozen tissue sections.					
Usage	All these reagents are effectively formulated for direct immunofluorescent staining of human tissue for flow cytometric analysis using 10 μ l/10 ⁶ leukocytes for singles and 20 μ l/10 ⁶ leukocytes in case of dual and triple combinations. Since applications vary, each investigator should titrate the reagent to obtain optimal results.					
HLDA Worksh	4 th Leukocyte Typing Workshop - Knapp, B., et al Eds. , Oxford University Press (1989).					
Representativ	/e Data					

Clone DF1524 (CD11a) was analyzed by flow cytometry using normal blood leucocytes. Direct staining was performed using 10 μ l of R-PE-conjugated antibody and 100 μ l blood sample.



IQP-151 – CD11a (DF1524)

Limitations

- 1. Conjugates with brighter fluorochromes, like PE and APC, will have a greater separation than those with dyes like FITC and CyQ. When populations overlap, the percentage of positive cells using a selected marker can be affected by the choice of fluorescent label.
- 2. Use of monoclonal antibodies in patient treatment can interfere with antigen target recognition by this reagent. This should be taken into account when samples are analyzed from patients treated in this fashion. IQ Products has not characterized the effect of the presence of therapeutic antibodies on the performance of this reagent.
- 3. Reagents can be used in different combinations, therefore laboratories need to become familiar performance characteristics of each antibody in relation with the combined markers in normal and abnormal samples.
- 4. Reagent data performance is based on EDTA-treated blood. Reagent performance can be affected by the use of other anticoagulants.

Reagents and materials required but not supplied

- 1. Flow cytometer
- 2. Flow cytometry disposable 12 x 75-mm capped polystyrene test tubes
- 3. Micropipette with disposable tips
- 4. Vortex mixer
- 5. Centrifuge
- 6. IQ Lyse erythrocyte lysing solution (IQP-199)
- 7. IO Starfigs fixation and permeabilization solution (IOP-200)
- 8. PBS (phosphate-buffered saline)
- 9. 1% paraformaldehyde solution in PBS (store at 2-8 °C in amber glass for up to 1 week)

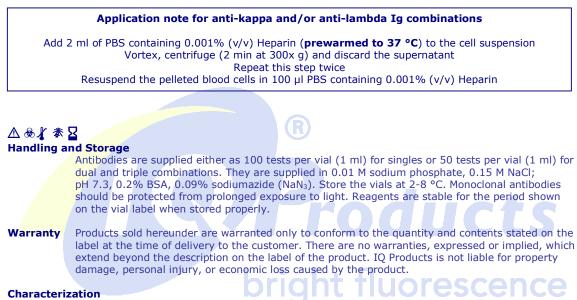
Immunofluorescence staining and lysing protocol

Flow cytometry method for use with purified monoclonal antibodies

- 1. Add 100 µl of EDTA-treated blood (i.e. approx. 10⁶ leukocytes) to a 5 ml reagent tube. The content of one tube is sufficient to perform one test.
- 2. Add to each tube 10 µl of purified monoclonal antibody*. Vortex the tube to ensure thorough mixing of antibody and cells.
- 3. Incubate the tube for 15 minutes at room temperature in the dark.
- Wash the labeled cells by adding 2 ml of PBS containing 0.001% ($^{v}/_{v}$) Heparin, vortexing and centrifuging 4. (2 min 1000 x g.) and discard the supernatant.
- Add 50 µl of 1:10 dilution of IQ Products F(ab)2 Rabbit Anti Mouse IgG fluorescent conjugate, [FITC (IOP-5. 190F); R-PE (IQP-190R)] in PBS containing 0.001% ($^{\nu}/_{\nu}$) Heparin to the tube. It is recommended that the tube is protected from light.
- Mix by vortexing and incubate for 15 minutes at room temperature in the dark. 6.
- Add 100 µl of IQ Lyse (IQP-199 ready-to-use) and mix immediately. 7.
- 8. Incubate for 10 minutes at room temperature in the dark.
- Add 2 ml of demineralized water and incubate for 10 minutes in the dark. 9.
- Centrifuge the labeled cell suspension for 2 minutes at 1000 x g. 10.
- Remove the supernatant and resuspend the cells in 200 µl of PBS**. 11.
- Analyze by flow cytometry within four hours (alternatively, the cells may be fixed by 0.05% of formaline 12. in buffered saline for analysis the next day. Some antigens are readily destroyed upon fixation and this should be taken into account when using this alternative).
- <u>- B</u> -Flow cytometry method for use with labeled (FITC, R-PE, Cy-Q or APC) monoclonal antibodies
- 1 Add 100 µl of EDTA-treated blood (i.e. approx. 10⁶ leukocytes) to a 5 ml reagent tube. The content of one tube is sufficient to perform one test.
- 2. Add to each tube 10 µl of labeled monoclonal antibody*. Vortex the tube to ensure thorough mixing of antibody and cells.
- 3. Incubate the tube for 15 minutes at room temperature in the dark.
- 4. Add 100 µl of IQ Lyse (IQP-199 ready-to-use) and mix immediately.
- Incubate for 10 minutes at room temperature in the dark. 5.
- 6. Add 2 ml of demineralized water and incubate for 10 minutes in the dark.
- Centrifuge the labeled cell suspension for 2 minutes at 1000 x g. 7.
- Remove the supernatant and resuspend the cells in 200 μ l of PBS**. 8.
- Analyze by flow cytometry within four hours (alternatively, the cells may be fixed by 0.05% of formaline 9 in buffered saline for analysis the next day. Some antigens are readily destroyed upon fixation and this should be taken into account when using this alternative).

- *C* Flow cytometry method for use with dual and triple combinations
- 1. Add 100 μ l of EDTA-treated blood (i.e. approx. 10⁶ leukocytes) to a 5 ml reagent tube. The content of one tube is sufficient to perform one test.
 - For combinations with anti-kappa and/or anti-lambda Ig see application note below.
- 2. Add to each tube 20 µl of labeled monoclonal antibody combination*.
- 3. Vortex the tube to ensure thorough mixing of antibody and cells.
- 4. Incubate the tube for 15 minutes at room temperature in the dark.
- 5. Add 100 μI of IQ Lyse (IQP-199 ready-to-use) and mix immediately.
- 6. Incubate for 10 minutes at room temperature in the dark.
- 7. Add 2 ml of demineralized water and incubate for 10 minutes in the dark.
- 8. Centrifuge the labeled cell suspension for 2 minutes at 1000 x g.
- 9. Remove the supernatant and resuspend the cells in 200 µl of PBS**.
- 10. Analyze by flow cytometry within four hours (alternatively, the cells may be fixed by 0.05% of formaline in buffered saline for analysis the next day. Some antigens are readily destroyed upon fixation and this should be taken into account when using this alternative).

* Appropriate mouse Ig isotype control samples should always be included in any labeling study ** PBS: Phosphate Buffered Saline, pH 7.2



To ensure consistently high-quality reagents, each batch of monoclonal antibody is tested for conformance with characteristics of a standard reagent. Representative flow cytometric data is included in this data sheet.

Warning All products contain sodiumazide. This chemical is poisonous and hazardous. Handling should be done by trained staff only.

References

- 1. Barclay, AN., 1997, The Leucocyte Antigen FactsBook. Academic Press. London. 156-157
- 2. Pigott, R., Power, C., 1993. The adhesion Molecule FactsBook. Academic Press, London
- 3. Lub, M et al., 1995. Immunol.Today, 16. 479-483
- 4. Wright, S.D., and Jong, M.T.C., 1986. J. Exp. Med., 164. 1876 1888
- 5. Knapp, B., et al eds. 1989 Leucocyte Typing IV. Oxford University Press

Explanation of used symbols

	Consult instructions for use				
REF	Catalogue number				
$\overline{\mathbb{V}}$	Sufficient for				
IVD	In Vitro Diagnostic medical device				
$\overline{\mathbb{A}}$	Caution, consult accompanying document				
*	Keep away from (sun)light				
&	Biological risks				
*	Temperature limitation (°C)				
RUO	For Research Use Only				
LOT	Batch code				
2	Use by yyyy-mm-dd				
	Manufacturer				
EC REP	Authorized Representative in the European Community				
CE	Conformité Européenne (European Conformity)				

		Label - tandem	Ex -max (nm)	Em -max (nm)
Р	PURE	purified material	-	-
F	FITC	FITC	488	519
R	R-PE	PE	488, 532	578
С	CyQ	PE-Cy5.18	488, 532	667
Α	APC		595, 633, 635, 647	660
PC	PerCP		488, 532	678
PCC	PerCP-Cy5.5		488, 532	695

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